



Deletion of the angiopoietin receptor Tie2 enhances proliferation and sprouting of cardiac endothelial cells

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Abstract

Background Endothelial cells (ECs) of the heart proliferate and form new vessels in response to vascular endothelial growth factor (VEGF), but VEGF has not benefited the therapy of cardiac ischemia because of its side effects. Here, we explored if deletion of the vascular steady-state homeostasis maintaining *Tie1* and *Tie2* receptor tyrosine kinases affects the proliferation and sprouting of cardiac ECs.

Methods We analyzed EC proliferation and histological and immunohistochemical stainings by confocal microscopy, plus scRNA and qPCR analyses of gene expression in the heart, kidneys, and lungs of *Tie1*^{fl/fl}, *Tie2*^{fl/fl}, and *Tie1*^{fl/fl}; *Tie2*^{fl/fl} mice, in which vascular endothelial cadherin-driven *CreER*^{T2} recombinase was used to delete *Tie1*, *Tie2* or both receptors. These analyses were also performed in mice subjected to transverse aortic constriction (TAC). Boyden chamber assays were performed to assess the migration of cultured ECs in cultures with or without *TIE* receptor silencing.

Results Genetic deletion of *Tie1*, *Tie2*, or *Tie1/Tie2* in mice increased significantly the proliferation of cardiac but not renal or pulmonary ECs, as measured by EdU incorporation into DNA and quantification of the cell cycle marker cyclin D1. *Tie1/Tie2* or *Tie2* deletion, but not *Tie1* deletion alone, induced EC sprouting in coronary vasculature and expression of endothelial tip cell markers, including expression of the FOXO1-regulated *Angpt2* and *Esm1* genes in cardiac versus kidney or lung ECs. Consistent with these findings, silencing of *TIE2*, but not *TIE1*, in cultured ECs resulted in increased migration of ECs. Similar results were obtained in mice subjected to TAC.

Conclusion Deletion of *Tie2* alone or together with *Tie1* increases the proliferation and sprouting of cardiac, but not renal or pulmonary ECs, without to neovessel formation in the heart.

Keywords Proliferation · Sprouting · Tie2 receptor · Angpt2 · Tip cell markers

Introduction

Cardiovascular diseases are the leading cause of mortality worldwide [1]. Although several treatments are available for primary and secondary prevention of atherosclerosis-associated coronary artery disease, insufficient blood supply to cardiac muscle often necessitates invasive restoration of coronary blood flow [2]. Therefore, alternative methods are continuously sought for treatment of myocardial ischemia. It has been shown that in acute myocardial infarction, clonal expansion of endothelial cells (ECs) occurs in the heart as part of an often insufficient neovascularization process [3]. Cardiac delivery of vascular growth factors via gene therapy has been tried for stimulation of neovascularization in the ischemic heart, but without clinical success [4].

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ECs regulate vascular permeability and leukocyte transmigration at intercellular junctions formed by several transmembrane proteins [5]. The TIE1 and TIE2 (also known as TEK) receptor tyrosine kinases form heteromeric complexes that are connected *in trans* across the junctions of neighboring ECs through their angiopoietin (ANGPT) ligands [6–8]. ANGPT1 stabilizes multimeric TIE2/TIE1 complexes, which is an essential mechanism for the maintenance of stable intercellular junctions [6, 7], whereas defective inter-endothelial junctions and loss of pericytes contribute to vessel destabilization [9, 10]. In response to challenges, such as disturbed blood flow or inflammation, TIE2 phosphorylation in ECs is decreased by autocrine expression of ANGPT2, which is a weaker agonist than ANGPT1, and by the endothelial phosphatase VE-PTP [11–13]. Constitutive deletion of *Tie2* impairs vascular stability during embryonic development [14, 15]. Interestingly, cardiac-specific deletion of *Angpt1* was sufficient to phenocopy the *Tie2* deletion phenotype in embryos, suggesting that continuous TIE2 activation is required for vascular stabilization [16, 17]. Furthermore, induction of autocrine ANGPT2 expression in ECs acts as an antagonist of the ANGPT1-TIE2 signaling to induce vessel destabilization during development and in hypoxic or inflamed tissues [12, 18, 19]. Thus, constitutive ANGPT1-dependent activation of TIE2 is important for vascular stabilization. Consistent with this, heterozygous *Tie2*-targeted mice show severe inflammatory stress, vascular leakage, and mortality [20, 21].

Evidence published so far suggests that the loss of *Tie2* expression and thus loss of TIE2 signals, caused by genetic deletion, gene silencing, or during various pathological stresses leads to altered gene expression and phenotypic response, such as attenuation of endothelial barrier function and induced vascular leakage [21]. In the present study, we aimed to further understand organ-specific cardiac functions of *Tie2* and *Tie1* and their possible connection with the poor angiogenic potential of cardiac ECs [22]. We used complete ablation of TIE signaling in endothelium by deleting either *Tie1* or *Tie2* receptor, or both receptors simultaneously in homeostatic or ischemic conditions caused by transverse aortic constriction (TAC), and analyzed EC proliferation, sprouting and vessel growth in the heart, kidneys, and lungs.

Materials and methods

Detailed experimental procedures are available in the Supplemental Materials and Methods.

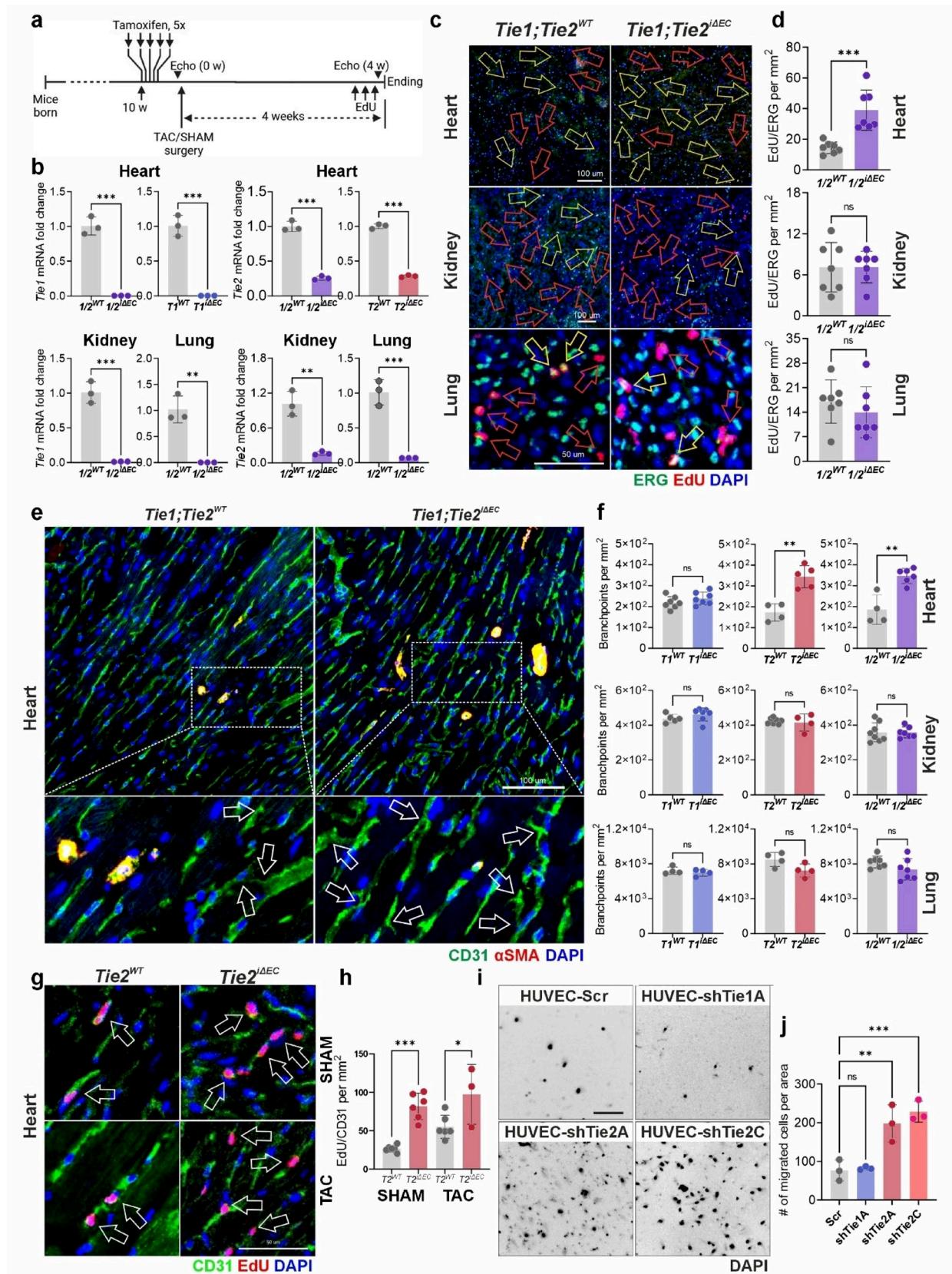
Fig. 1 Deletion of *Tie1* and *Tie2* alters proliferation and sprouting of cardiac endothelial cells. **a** Time course of the experiment. **b** qRT-PCR analysis of *Tie1* and/or *Tie2* mRNA levels in total lysates from the target tissues. Marking here onwards: $1/2^{WT}$ and $1/2^{iAEC} = Tie1; Tie2^{WT}$ and $Tie1; Tie2^{iAEC}$, respectively; $T1^{iAEC} = Tie1^{iAEC}$, $T2^{iAEC} = Tie2^{iAEC}$ ($n=3$ per group). **c** Representative images of immunofluorescently stained sections from the target organs for EdU and ERG. Yellow-outlined arrows indicate EdU^+/ERG^+ nuclei; red-outlined arrows indicate EdU^+/ERG^- nuclei. Only images from $Tie1; Tie2^{iAEC}$ (vs. control) groups are shown. **d** Quantifications of (c). Quantifications of EdU/ERG from the target organs of *Tie1*- or *Tie2*-deleted (vs. control) groups are shown in Fig. S1e ($n=7$ per group). **e** Representative images of sections from heart immunofluorescently stained for CD31 and α SMA. Arrows indicate examples of the branch points (described in the Materials and methods). Representative images of CD31/ α SMA-stained sections of kidney and lung are shown in Fig. S1f and g. **f** Quantification for branchpoints from the images in (e) and in Fig. S1f and g from heart, kidney and lung ($n=4$ –8 per group). **g** and **h** Representative images and quantification of cardiac sections from the mice four weeks after SHAM or TAC operations ($n=3$ –6 per group). **i** Experiment using Boyden chamber and HUVECs, silenced for *TIE1* or *TIE2* using lentivirus-delivered shRNA. Representative images of the membranes showing transmigrated DAPI-stained cells on the lower side of the membrane. **j** Quantification of (i). Scale bars in (e) heart and kidney 100 μ m; lung 50 μ m; in e=100 μ m; in g 50 μ m ($n=3$ per group). For statistics in the graphs in (b), (d) and (f) a 2-tailed Student's *t*-test was used. For statistics on graphs (h) and (j) one-way ANOVA followed by Dunnett's *post hoc* test was used. Each dot represents average value per mouse. * $P<0.05$; ** $P<0.01$; *** $P<0.001$

Results

Deletion of *Tie2* or *Tie1/Tie2* increases endothelial cell proliferation and network complexity in cardiac, but not renal or pulmonary vasculature

To analyze the importance of TIE receptor signals in cardiac blood capillaries, we generated mice carrying *Cdh5-BAC-CreER^{T2}* transgene plus conditionally targeted *Tie1^{f/f}*, *Tie2^{f/f}* alleles or their combination (Fig. S1a). We then deleted the Tie receptors from ECs under homeostatic conditions by administering tamoxifen to 10-week-old male mice. Efficient deletion of both *Tie1* and *Tie2* encoding mRNAs, without deletion of the intervening region in chromosomal DNA, was confirmed by qPCR (Figs. 1a, b and S1b, c and [23]). We also confirmed the lack of compensation of the deleted receptor *Tie1* or *Tie2* from the partnering receptor *Tie2* or *Tie1*, respectively (Fig. S1d).

To analyze the proliferation effects of *Tie* deletion 4 weeks after tamoxifen administration, mice with an EC-specific deletion of only *Tie1* (*Tie1^{iAEC}*), only *Tie2* (*Tie2^{iAEC}*), or both *Tie1* and *Tie2* deletions (*Tie1; Tie2^{iAEC}*), were injected with six doses of EdU during three consecutive days and



terminated 12 h after the last EdU dose. Immunofluorescence stainings and analysis of tissue sections showed that both single and compound deletions led to a significant increase of DNA synthesis in cardiac but not renal or pulmonary ECs, as measured by EdU incorporation into DNA (Figs. 1c, d, S1e). In the hearts of either control or *Tie*-deleted mice, EdU was found to be relatively evenly distributed without accumulation close to the pericardium or the endocardium.

The cardiac vascular network in gene-deleted and control mice, analyzed by quantification of the number of branch points per unit of imaging area, revealed an increase in vascular complexity in the hearts of the *Tie2*^{iAEC} and *Tie1; Tie2*^{iAEC} mice, whereas the difference was not significant in *Tie1*^{iAEC} mice ($p=0.2362$). There was no significant difference in branch points when comparing gene-deleted kidney or lung in any of the three genotypes analyzed (Figs. 1e, f and S1f, g). The overall vessel-covered area in the cardiac sections was not increased (Fig. S1h). There was no increase in vascular area percentage in renal or pulmonary vessels after deletion of *Tie1*, *Tie2*, or both (Figs. 1f and S1f–h). Furthermore, a small but significant decrease in the vessel-covered area was observed in the lungs of *Tie1; Tie2*^{iAEC} mice (Fig. S1h).

We next analyzed if the effect of *Tie2* deletion in the heart is affected by ischemic conditions induced by TAC, which reduces the diameter of the aorta, leading to pressure overload and adverse cardiac remodeling. *Tie2*^{iAEC} mice and control littermates were treated with tamoxifen and subjected to aortic constriction. Four weeks later, the mice were injected with EdU (twice per day during the last 3 days) and euthanized 12 h thereafter. Analysis of EdU counts associated with CD31-positive ECs revealed an approximately two-fold increase, plus increased branchpoint formation in mice that underwent either SHAM or TAC surgery, but again, there was no significant increase in the overall vessel density (Figs. 1g, h and S2a, b). In echocardiography of the SHAM and TAC groups, *Tie2* deletion did not seem to have any effect. In homeostatic conditions, *Tie1*^{iAEC} did not affect the echocardiography parameters. *Tie2*^{iAEC} affected one echocardiographic parameter, whereas *Tie1; Tie2*^{iAEC} affected five parameters, yet none of the groups, including the TAC group, showed significant differences in ejection fraction (EF) or fractional shortening (FS), indicating no significant changes in myocardial function (Supplemental Table 1). *Tie1* and *Tie2* deletions did not cause hypertrophy or fibrosis in the myocardium (Figs. S2d–f and S3a–c). Furthermore, CD45+ cell accumulation was similar between *Tie1/Tie2*-deleted and control mice, indicating no increase in cardiac inflammation (Fig. S3d–f).

Since vessel branch points develop mainly via vessel sprouting and EC migration in adult mice, we next analyzed

if the silencing of *TIE1* or *TIE2* mRNA and the respective proteins affects the migration of cultured ECs. Using a Boyden chamber assay, we tested the migration capacity of human umbilical vein ECs (HUVECs) after silencing *TIE1* or *TIE2* mRNA via lentiviral shRNAs (Fig. S4a). We found that *TIE2*, but not *TIE1* silencing, led to a significant increase in EC migration rate (Fig. 1i, j), which was consistent with the analysis of RNA profiles by hierarchical clustering in HUVECs after *TIE2*, but not *TIE1* silencing (Fig. S4b, c). Furthermore, a striking morphological and kinetic difference was observed between the *shTIE1*, *shTIE2*, and control vector-transfected cells in an EC monolayer wound closure assay. The most conspicuous difference between the two was the elongation of the ECs, accompanied by their improved migration through the 8 μ m pores of the Boyden chamber membranes after *TIE2*, but not *TIE1*, silencing (Supplemental videos S1–S3 and Fig. S4d for quantification).

These data indicated that loss of *TIE1/TIE2* or *TIE2* in mice results in increased proliferation and sprouting of cardiac, but not renal or pulmonary ECs in homeostatic conditions and under ischemic cardiac stress in the TAC model. *TIE2* silencing seemed to increase EC elongation and migration more than *TIE1* silencing in cultured ECs.

Enhanced expression of endothelial tip cell markers after *Tie1* and/or *Tie2* deletion

To analyze how the *Tie* deletion affected the EC transcriptome, we next performed single-cell RNA sequencing (scRNAseq) of EC-enriched fractions from the *Tie1*^{iAEC} vs. control *Tie1*^{WT} hearts and stromal-vascular fractions (SVF) of the *Tie2*^{iAEC} and *Tie1; Tie2*^{iAEC} versus their corresponding controls (*Tie2*^{WT} and *Tie1; Tie2*^{WT}, respectively). Figure 2a and b shows the scRNAseq workflow of the cardiac SVF samples after four weeks of gene deletion. We identified 19 distinct cell clusters in cardiac SVF from the *Tie1; Tie2*^{iAEC} mice (Supplemental Table 2). Based on low expression of most genes, damaged cells were excluded, leaving 16 clusters for further analyses (Fig. 2c). Of these, six were identified as endothelial clusters based on commonly known EC markers, such as *Flt1*, *Kdr*, *Cldn5*, and *Pecam1* for blood vascular ECs (BECs), and *Prox1*, *Ccl21a*, and *Flt4* for lymphatic ECs (LECs). The BEC subtypes were further sub-classified as capillary-like EC I (*Aplnr* and *Car4*), arterial (*Hey1* and *Cxcl12*), activated (*Kit* and *Apln*), venous (*Vwf* and *Vcam1*), and *Ifit*+ ECs (*Ifit1* and *Ifit3*) (Fig. S5a, Supplemental Table 2). In line with the immunohistochemical data, scRNAseq analyses confirmed that both single and compound deletions of the Tie receptors increased the number of proliferating cardiac ECs (Figs. 2c and S5b, c [figure legend for the cell numbers]; Supplemental Table 2). We also confirmed that the majority of the proliferating cells

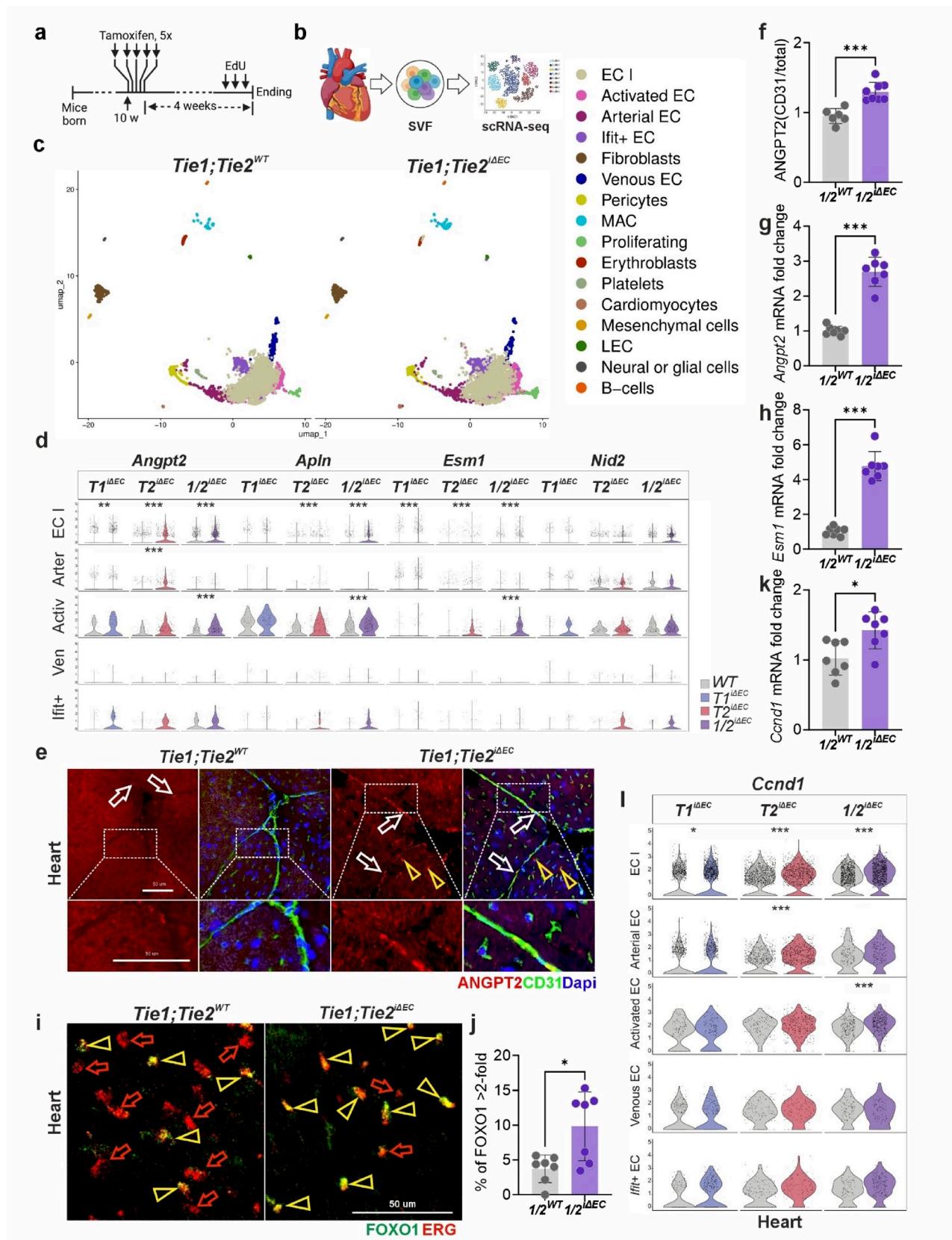


Fig. 2 Expression of tip cell markers in cardiac endothelial cells. **a** Time course of the scRNAseq experiment. **b** Schematic of the tissue processing for scRNAseq analysis. **c** UMAP plots (subsampled to 3580 cells each), showing cluster distribution in SVF from the hearts of *Tie1*/*Tie2*^{WT} and *Tie1*/*Tie2*^{iAEC} mice. Cluster names are shown on the right. The numbers of *Mki67*⁺*ERG*⁺ cells in *Tie1*/*Tie2*^{WT} and in *Tie1*/*Tie2*^{iAEC} samples are 23 and 72, respectively. Analysis parameters and cluster markers are shown in Supplemental Tables 4 and 5, respectively. More data regarding differentially expressed genes and associated pathway analysis for this tissue and genetic model can be found in Supplemental Tables 10, 15 and 16. For gene signatures of the five major endothelial clusters see Fig. S5a; for UMAP plots of *Tie1*^{WT} vs. *Tie1*^{iAEC} and *Tie2*^{WT} vs. *Tie2*^{iAEC} hearts see Fig. S5b and c. **d** Violin plots showing relative expression of the four tip cell marker genes *Angpt2*, *Apln*, *Esm1* and *Nid2* in five major EC clusters from the hearts of three genotypes *Tie1*/*Tie2*^{iAEC} (1/2^{iAEC}), *Tie2*^{iAEC} (2^{iAEC}) and *Tie1*^{iAEC} (T1^{iAEC}). Pairwise comparisons are made with the corresponding controls (grey). Violin plots are given the same scale of [0, 5]. Asterisks indicate a significant difference between the neighboring violin plots, i.e., the wild-type control (grey) and the *Tie*-deleted one (colored). **e** and **f** Representative images and quantification of ANGPT2 immunofluorescence staining of the heart sections from *Tie1*/*Tie2*^{WT} vs. *Tie1*/*Tie2*^{iAEC} mice. Quantification represents the ratio of ANGPT2⁺ fluorescence intensity (F.I.) within the CD31⁺ areas (calculated as F.I. per μm^2), divided by the “total” ANGPT2 F.I. from the whole image area (also calculated as F.I. per μm^2) ($n=6$ –8 per group). **g** and **h** qRT-PCR analysis of *Angpt2* and *Esm1* mRNA expression in the whole heart lysates of *Tie1*/*Tie2*^{WT} vs. *Tie1*/*Tie2*^{iAEC} mice ($n=7$ per group). **i** and **j** Representative images and quantification of cardiac sections immunofluorescently stained for FOXO1 and ERG in *Tie1*/*Tie2*^{WT} vs. *Tie1*/*Tie2*^{iAEC} mice. See Supplemental Materials and Methods section for the details of the quantification ($n=7$ per group). **k** qRT-PCR analysis of *Ccnd1* mRNA expression in the total heart lysates of *Tie1*/*Tie2*^{WT} vs. *Tie1*/*Tie2*^{iAEC} mice ($n=7$ per group). **l** Violin plots showing relative expression of the *Ccnd1* mRNA in five major EC clusters from the hearts of three genotypes *Tie1*/*Tie2*^{iAEC}, *Tie2*^{iAEC} and *Tie1*^{iAEC}. Scale bars are 50 μm . Pairwise comparisons are made with the respective undeleted controls, shown on the graph as violins colored in grey. For scRNAseq data, Wilcoxon Rank Sum test was used. For statistics in the graphs in (f)–(h), (j) and (k) 2-tailed Student’s *t*-test was used; a Welch’s correction was used for (j). Each dot represents average value per mouse. * $P<0.05$; ** $P<0.01$; *** $P<0.001$

in the heart in our genetic models were ECs, as determined by co-expression of *Mki67* and *Erg* (ETS-Related Gene) markers in our scRNAseq data (Supplemental Table 3). All parameters used for quality control, filtering, normalization, integration and clustering are shown in Supplemental Table 4. Cluster markers for all analyzed genotypes and tissues are listed in Supplemental Tables 5–9.

As sprouting correlates with the expression of tip cell markers, we next investigated if the deletion of *Tie1* and/or *Tie2* genes stimulated the expression of well-known tip cell marker genes, such as *Angpt2*, *Apln*, *Esm1*, and *Nid2* [24, 25]. Indeed, we found a significant upregulation of the tip cell markers, especially in cardiac ECs of the capillary-like cluster, in the *Tie1*/*Tie2*^{iAEC} and *Tie2*^{iAEC} mice, and to a lesser extent in the *Tie1*^{iAEC} mice (Fig. 2d). Other differentially expressed genes for all analyzed genotypes and tissues are listed in Supplemental Tables 10–14. Immunofluorescence staining was used to confirm increased expression

of ANGPT2 in the cardiac sections from the *Tie1*/*Tie2*^{iAEC} mice (Fig. 2e, f). Furthermore, qRT-PCR analysis of total heart lysates confirmed increased expression of *Angpt2* and *Esm1* RNAs in the *Tie1*^{iAEC}, *Tie2*^{iAEC}, and *Tie1*/*Tie2*^{iAEC} mice vs. undeleted control mice (Fig. 2g, h and S5d, e), as well as in the SHAM and TAC-treated mice (Fig. S2c).

The scRNAseq and qRT-PCR analyses of tip cell markers in renal ECs showed *Esm1* upregulation in response to *Tie2* deletion, but only a weak expression of *Angpt2* and *Apln* (Fig. S6a–e). *Tie1* deletion did not lead to upregulation of *Angpt2* or *Esm1* RNA (Fig. S6d, e). Analysis of pulmonary ECs revealed a small, but significant upregulation of *Esm1* RNA in the *Tie1*/*Tie2*^{iAEC} mice. scRNAseq analysis furthermore revealed no significant alterations of the four tip cell marker genes (Fig. S7a–e). Thus, our results of the tip cell marker gene expression are consistent with the findings of increased branch points in cardiac, but not pulmonary or renal vessels.

Increased nuclear localization of FOXO1 and ERG in *Tie*-deleted cardiac but not renal or pulmonary ECs

The FOXO1 and ERG transcription factors are abundantly expressed in ECs [26, 27]. Upon activation, they tend to accumulate in the nucleus where they transactivate their target genes. Ligand stimulation of TIE1/TIE2 activates the AKT kinase, which phosphorylates FOXO1, leading to its nuclear exclusion [11, 12]. We found a significant increase of FOXO1 signal in the nuclei of cardiac ECs of the *Tie1*/*Tie2*^{iAEC} mice (Fig. 2i, j), which is consistent with the upregulation of tip cell marker genes (Fig. 2d–h), including *Angpt2*, *Esm1*, which are known FOXO1 target genes [24]. In contrast, no increase of nuclear FOXO1 was observed in *Tie1*/*Tie2*-deleted pulmonary ECs (Fig. S8a, b). ERG was markedly accumulated in the nuclei of *Tie1*/*Tie2*- and to a lesser extent in *Tie2*-deleted cardiac ECs. No increase of nuclear ERG was observed in ECs of *Tie*-deleted kidneys or lungs (Fig. S8c–f).

Transcripts encoding the main G₀–G₁ switcher cyclin D1 (*Ccnd1*) were strongly upregulated in major cardiac EC clusters (EC I, Arterial EC and Activated EC; scRNAseq data) of *Tie1*/*Tie2*^{iAEC} and *Tie2*^{iAEC} mice and in total *Tie1*/*Tie2*^{iAEC} cardiac mRNA by qRT-PCR (Figs. 2k, l and S9a). Consistent with the EdU staining results, the expression of *Ccnd1* was not significantly altered in any EC sub-clusters of kidney or lung RNA in the scRNAseq data (Figs. S6f and S7f). As *Ccnd1* is also a direct FOXO1 target [28], its upregulation suggests that the increase in cell proliferation could partly be mediated by FOXO1. We also found that cyclin-dependent kinase 8 (*Cdk8*), involved in cell cycle progression, was strongly upregulated upon *Tie1*/*Tie2* and *Tie2* deletion in cardiac, but not renal or pulmonary ECs

(Figs. S6f, S7f and S9b). The results suggest that, unlike the ECs in the kidney and lung, cardiac ECs remain sensitive to TIE signals, in part, via regulation of the intracellular localization of ERG and FOXO1 in ECs.

Discussion

Our recent studies on the role of the TIE1 and TIE2 receptor signaling in atherosclerosis [23] led us to ask how deletion of the Tie receptor signaling system in mice under homeostatic conditions affects coronary vessels. We show here that, unlike aortic ECs, loss of TIE2 or both TIE1 and TIE2, but not TIE1 alone, in cardiac ECs promotes endothelial proliferation and sprouting. This was an organotypic response that was not observed in the kidneys or the lungs. Surprisingly, also *Tie2* deletion in TAC yielded results corresponding to *Tie2* deletion in homeostatic conditions.

We demonstrated a strong upregulation of *Angpt2* mRNA and ANGPT2 protein expression in the heart upon Tie deletions. As this response was lacking in the kidneys or lungs, it seems to be an organotypic response in cardiac ECs that may depend on specific activation of transcription factors, such as FOXO1 and ETS-1, which both bind to the promoter of the *Angpt2* gene to activate *Angpt2* expression [24, 29]. ANGPT2 in *Tie2*-deleted ECs can bind to and activate $\alpha 5\beta 1$ -integrin leading to increased phosphorylation of mitogen-activated protein kinase (MAPK) ERK1/2 [30], which is a master regulator of the G1- to S-phase transition [31]. We show that the GO terms related to ERK1, ERK2 and the MAPK cascade were significantly upregulated upon *Tie1*/ *Tie2*- or *Tie2*-deletion or silencing in cardiac ECs or cultured ECs, respectively (Figs. S4b, c and S9c, d). Other GO terms for all analyzed genotypes and tissues are listed in Supplemental Tables 15–24. Likewise, the lack of *Angpt2* upregulation in renal and pulmonary ECs in response to *Tie* deletion (as we show here) correlated with the lack of ECs' proliferation response in these two organs.

The key endothelial transcription factor ERG was also upregulated in the *Tie1/Tie2* and *Tie2*-deleted cardiac, but not in renal or pulmonary ECs. ERG plays a crucial role in promoting angiogenesis and vascular stability during development and also postnatally. In mature vasculature, ERG also functions to maintain endothelial homeostasis [27]. The increased ERG expression could contribute also to the vascular branching that we observed, and the upregulation of the FOXO1 regulated *Ccnd1* in the *Tie*-deleted cardiac ECs could contribute to increased EC proliferation [28].

It should be noted that our results are consistent with the view that TIE1, which is a weaker tyrosine kinase than TIE2, mainly stabilizes and enhances TIE2 signaling rather than representing a standalone signaling receptor [7, 32].

TIE1 and TIE2 receptors in their transmembrane complex are active in signaling via the PI3-kinase-AKT pathway in steady-state homeostatic ECs of blood vessels [32, 33]. The PI3K-AKT pathway leads to phosphorylation of downstream molecular targets, including the FOXO1 transcription factor, which is thereby sequestered in the cytoplasm where it can be degraded via polyubiquitylation and proteasomal degradation [34]. Unphosphorylated FOXO1 is retained in the nucleus and transactivates target genes such as *Angpt2*, *Esm1* and *Ccnd1* via binding to the promoter/enhancer regions of these genes [24], contributing to the angiogenic events, as shown in this manuscript.

Interestingly, the expression of the tip cell markers *Angpt2*, *Esm1* and *Ccnd1* upon *Tie1*, *Tie2* or *Tie1/Tie2* deletion was low or undetectable in the kidney and lung, respectively. The attenuated response in renal and pulmonary ECs could result from cell- and tissue-type specificity of FOXO1 binding and activation. Specifically, FOXO1 activates a distinct sets of genes in different cell types, as has been shown for ECs and B-cells using chromatin immunoprecipitation (ChIP) analysis. The transcriptional specificity was mediated via the co-binding of ETS and GATA transcription factors in ECs, and ETS and IRF in B-cells [24]. Thus, the intracellular genomic microenvironment in cardiac versus renal and pulmonary ECs could be responsible for the tissue specificity of the FOXO1-mediated transcriptional responses, which could be further investigated in future studies.

It is noteworthy that while we observed increased EdU incorporation following *Tie* deletion in the heart, this did not translate into an overall increase of the cardiac vasculature. Thus, our genetic model did not display a complete angiogenic response. Some angiogenesis-related responses were present, such as enhanced branching, yet we did not detect capillary elongation or fusion. This may not be surprising given that even the master regulator of angiogenesis, VEGF, exhibits a limited efficacy in the heart. For example, in the study by Kocjan et al., VEGF overexpression in skeletal muscle led to the formation of a large number of new capillaries and arterioles. In contrast, the response to the same dose of VEGF in the heart was blunted, yielding only a modest increase in new arterioles [22], suggesting that intrinsic mechanisms within the heart may regulate the angiogenic response.

In conclusion, we discovered that deletion of Tie2 induces cardiac endothelial proliferation and sprouting without vessel formation in the heart. This suggests that additional stimuli maybe ultimately required for angiogenesis. A schematic of our interpretation of the data is shown in Fig. S9e. Further studies should elucidate the therapeutic potential of manipulating the TIE1/TIE2 signaling in the context of therapeutic vascularization.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10456-025-10028-2>.

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Author contributions A.A., K.A. and S.K.—conceptualization and design of the study, analysis of the data, manuscript writing, figure preparation; A.A. and M.H.L.—data collection; H.G.A.—provision of the important study material; E.M.—provision of the necessary experimental technique (TAC surgery).

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Data availability Raw and filtered scRNAseq sequencing files are available on GEO (Accession Number GSE297297; <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE297297>). Other data are provided within the manuscript or supplementary material files.

Declarations

Conflict of interest The authors declare no conflict of interest.

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References

1. Lindstrom M, DeCleene N, Dorsey H et al (2022) Global burden of cardiovascular diseases and risks collaboration, 1990–2021. *J Am Coll Cardiol* 80:2372–2425. <https://doi.org/10.1016/j.jacc.2022.11.001>
2. Khalil M, Maqsood MH, Basir M et al (2022) TCT-7 invasive versus conservative strategy in elderly patients with non-ST-segment elevation myocardial infarction: a meta-analysis of randomized controlled trials. *J Am Coll Cardiol* 80:B3 <https://doi.org/10.1016/j.jacc.2022.08.015>
3. Manavski Y, Lucas T, Glaser SF et al (2018) Clonal expansion of endothelial cells contributes to ischemia-induced neovascularization. *Circ Res* 122:670–677. <https://doi.org/10.1161/CIRCRESAHA.117.312310>
4. Ferrara N, Alitalo K (1999) Clinical applications of angiogenic growth factors and their inhibitors. *Nat Med* 5:1359–1364
5. Dejana E, Orsenigo F (2013) Endothelial adherens junctions at a glance. *J Cell Sci* 126:2545–2549. <https://doi.org/10.1242/jcs.124529>
6. Fukuhara S, Sako K, Minami T et al (2008) Differential function of Tie2 at cell–cell contacts and cell–substratum contacts regulated by angiopoietin-1. *Nat Cell Biol* 10:513–526. <https://doi.org/10.1038/ncb1714>
7. Leppänen V-M, Saharinen P, Alitalo K (2017) Structural basis of Tie2 activation and Tie2/Tie1 heterodimerization. *Proc Natl Acad Sci USA* 114:4376–4381. <https://doi.org/10.1073/pnas.1616166114>
8. Saharinen P, Eklund L, Miettinen J et al (2008) Angiopoietins assemble distinct Tie2 signalling complexes in endothelial cell–cell and cell–matrix contacts. *Nat Cell Biol* 10:527–537. <https://doi.org/10.1038/ncb1715>
9. Komarova YA, Kruse K, Mehta D, Malik AB (2017) Protein interactions at endothelial junctions and signaling mechanisms regulating endothelial permeability. *Circ Res* 120:179–206. <https://doi.org/10.1161/CIRCRESAHA.116.306534>
10. Armulik A, Genové G, Mäe M et al (2010) Pericytes regulate the blood–brain barrier. *Nature* 468:557–561. <https://doi.org/10.1038/nature09522>
11. Carota IA, Kenig-Kozlovsky Y, Onay T et al (2019) Targeting VE-PTP phosphatase protects the kidney from diabetic injury. *J Exp Med* 216:936–949. <https://doi.org/10.1084/jem.20180009>
12. Saharinen P, Eklund L, Alitalo K (2017) Therapeutic targeting of the angiopoietin-TIE pathway. *Nat Rev Drug Discov* 16:635–661. <https://doi.org/10.1038/nrd.2016.278>
13. Thurston G, Daly C (2012) The complex role of angiopoietin-2 in the angiopoietin-tie signaling pathway. *Cold Spring Harb Perspect Med* 2:a006550. <https://doi.org/10.1101/cshperspect.a006550>
14. Dumont DJ, Gradwohl G, Fong GH et al (1994) Dominant-negative and targeted null mutations in the endothelial receptor tyrosine kinase, tek, reveal a critical role in vasculogenesis of the embryo. *Genes Dev* 8:1897–1909. <https://doi.org/10.1101/gad.8.16.1897>
15. Sato TN, Tozawa Y, Deutsch U et al (1995) Distinct roles of the receptor tyrosine kinases Tie-1 and Tie-2 in blood vessel formation. *Nature* 376:70–74. <https://doi.org/10.1038/376070a0>
16. Jeansson M, Gawlik A, Anderson G et al (2011) Angiopoietin-1 is essential in mouse vasculature during development and in response to injury. *J Clin Invest* 121:2278–2289. <https://doi.org/10.1172/JCI46322>

17. Suri C, Jones PF, Patan S et al (1996) Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. *Cell* 87:1171–1180. [https://doi.org/10.1016/s0092-8674\(00\)81813-9](https://doi.org/10.1016/s0092-8674(00)81813-9)
18. Gurnik S, Devraj K, Macas J et al (2016) Angiopoietin-2-induced blood-brain barrier compromise and increased stroke size are rescued by VE-PTP-dependent restoration of Tie2 signaling. *Acta Neuropathol* 131:753–773. <https://doi.org/10.1007/s00401-016-1551-3>
19. Maisonpierre PC, Suri C, Jones PF et al (1997) Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science* 277:55–60. <https://doi.org/10.1126/science.277.5322.55>
20. Ghosh CC, Mukherjee A, David S et al (2012) Impaired function of the Tie-2 receptor contributes to vascular leakage and lethality in anthrax. *Proc Natl Acad Sci USA* 109:10024–10029. <https://doi.org/10.1073/pnas.1120755109>
21. Ghosh CC, David S, Zhang R et al (2016) Gene control of tyrosine kinase TIE2 and vascular manifestations of infections. *Proc Natl Acad Sci USA* 113:2472–2477. <https://doi.org/10.1073/pnas.1519467113>
22. Kocjan T, Rehman M, Colliva A et al (2021) Genetic lineage tracing reveals poor angiogenic potential of cardiac endothelial cells. *Cardiovasc Res* 117:256–270. <https://doi.org/10.1093/cvr/vaa012>
23. Anisimov A, Fang S, Hemanthkumar KA et al (2023) The angiopoietin receptor Tie2 is atheroprotective in arterial endothelium. *Nat Cardiovasc Res* 2:307–321. <https://doi.org/10.1038/s44161-023-00224-y>
24. Miyamura Y, Kamei S, Matsuo M et al (2024) FOXO1 stimulates tip cell-enriched gene expression in endothelial cells. *iScience* 27:109161. <https://doi.org/10.1016/j.isci.2024.109161>
25. del Toro R, Prahst C, Mathivet T et al (2010) Identification and functional analysis of endothelial tip cell-enriched genes. *Blood* 116:4025–4033. <https://doi.org/10.1182/blood-2010-02-270819>
26. Potente M, Urbich C, Sasaki K et al (2005) Involvement of Foxo transcription factors in angiogenesis and postnatal neovascularization. *J Clin Investig* 115:2382–2392. <https://doi.org/10.1172/JCI23126>
27. Shah AV, Birdsey GM, Randi AM (2016) Regulation of endothelial homeostasis, vascular development and angiogenesis by the transcription factor ERG. *Vasc Pharmacol* 86:3–13
28. Yang L, Yang F, Zhao H et al (2019) Circular RNA circchfr facilitates the proliferation and migration of vascular smooth muscle via miR-370/FOXO1/Cyclin D1 pathway. *Mol Ther Nucleic Acids* 16:434–441. <https://doi.org/10.1016/j.omtn.2019.02.028>
29. Hasegawa Y, Abe M, Yamazaki T et al (2004) Transcriptional regulation of human angiopoietin-2 by transcription factor Ets-1. *Biochem Biophys Res Commun* 316:52–58 <https://doi.org/10.1016/j.bbrc.2004.02.019>
30. Hakanson L, Sipila T, Leppanen VM et al (2015) Endothelial destabilization by angiopoietin-2 via integrin β 1 activation. *Nat Commun* 6:5962 <https://doi.org/10.1038/ncomms6962>
31. Meloche S, Pouysségur J (2007) The ERK1/2 mitogen-activated protein kinase pathway as a master regulator of the G1- to S-phase transition. *Oncogene* 26:3227–3239
32. Korhonen EA, Lampinen A, Giri H et al (2016) Tie1 controls angiopoietin function in vascular remodeling and inflammation. *J Clin Investig* 126:3495–3510. <https://doi.org/10.1172/JCI84923>
33. Kim M, Allen B, Korhonen EA et al (2016) Opposing actions of angiopoietin-2 on Tie2 signaling and FOXO1 activation. *J Clin Investig* 126:3511–3525 <https://doi.org/10.1172/JCI84871>
34. Eijkelenboom A, Burgering BMT (2013) FOXOs: signalling integrators for homeostasis maintenance. *Nat Rev Mol Cell Biol* 14:83–97. <https://doi.org/10.1038/nrm3507>

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